Experimental elucidation of meiofaunal trophic interactions: From radioactive tracer techniques to next generation sequencing

Tom Moens, Nele De Meester, Katja Guilini, Anna-Maria Vafeiadou, Ann Vanreusel and Sofie Derycke



Elucidation of meiofaunal trophic interactions: Introduction

Meiofauna play also a key ecological role in linking detrital (and prokaryotic) resources with higher trophic levels: in fact most of the meiofaunal taxa eat microalgae, prokaryotes and detritus and, at the same time, it is known that meiofauna are a food source for macrofauna and fishes. Meiofauna and nematodes, based on laboratory and in situ experiments, are in fact able to influence microbial activities and to graze their production. *Pusceddu et al. (2014)*

Grazers can affect bacterial communities at different levels. They are able to influence bacterial activity, either stimulatory or inhibitory. This can be a direct effect of grazing, but bioturbation and secretion of mucus trails by nematodes can also be important. Grazing impacts on the bacterial community structure have also been reported.

De Mesel et al. (2004)

Meiofauna are ubiquitous in marine soft-sediment communities, and are an important link in transferring carbon primary and secondary production to higher trophic levels. Baguley et al. (2008)

Although their biomass is generally low, their high abundance and high metabolic and reproductive rates render them potentially

Important in benthic fluxes of carbon and nutrients (Kuipers et al., 1981, Coull, 1999). Moens et al. (2005)

Elucidation of meiofaunal trophic interactions Introduction

"Meiofauna matters: the roles of meiofauna in benthic

ecosystems

(Schratzberger & Ingels, keynote at this conference)

Why?

- non-trophic effects and interactions

- direct and indirect trophic interactions

"Meiofauna people are fond of arm-waving to make

speculations about how important meiofauna may be."

(anonymous reviewer, 2005)

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Elucidation of meiofaunal trophic interactions: Content

Approaches to measure and quantify direct trophic interactions

from past to future

methodological constraints/problems

Are nematodes quantitatively important consumers?

controversy between and across different approver

At what level should we measure (feeding types, families, species,

populations, individuals...)?

1. Observations

A. Some problems

- time consuming
- observations in sediments?
- \rightarrow under artificial conditions
- \rightarrow artificial food 'availability'
- → what set of 'environmental' conditions?
- 'snapshot' of reality \rightarrow anecdotal?
 - largely qualitative

B. Some common practices

- no observations
- observations in artificial media
 - \rightarrow under artificial conditions
 - \rightarrow artificial food availability
- \rightarrow most commonly at a constant

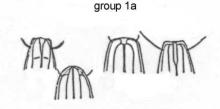
temperature (often room), in light,...

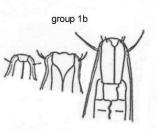
- observations of gut content →
 anecdotal and often inconclusive
- rely on morphological features

1. Observations

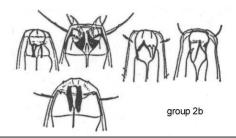
C. Some typical short-cuts

Wieser 1953

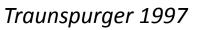








Black-box approach → 1
 species, 1 feeding type
 Similar species do the same





Moens & Vincx 1997

SIT FEEDERS EPIGROWTH FEEDE

Can't you see

I'm a vegetarian?!

1. Observations

C. Some typical short-cuts

- Feeding-type classifications are based more on how a nematode feeds than on what it eats

- Morphology can be misleading

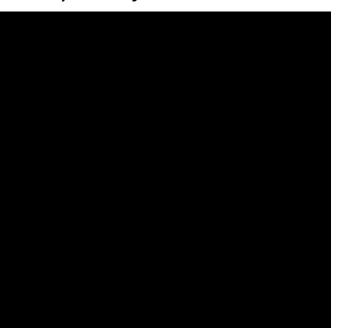
Hypodontolaimus & Metachromadora have a muscular pharynx and prominent tooth, but they are not predators

1. Observations

D. Some future directions

behavioral observations

courtesy An-Sofie D'Hondt



on artificial media, e.g. movement

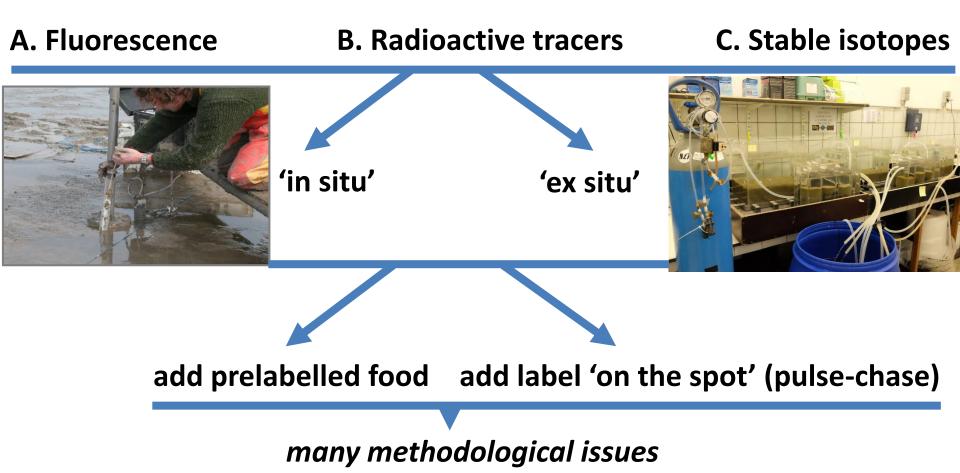
towards/selection of food

in the sediment matrix



courtesy Luana Monteiro

2. Tracer experiments



2. Tracer experiments

methodological issues

Use of prelabelled food

- choice of food (usually single species)
- alive, dead, preserved (how)?
- no realistic food distribution
- no realistic food-sediment 'interaction'

Pulse-chase

- not only the intended food can get labelled
- multiple non-grazing routes of label uptake
- how to properly administer and distribute label?

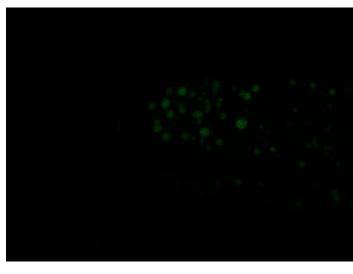
2. Tracer experiments

methodological issues in <u>fluorescent tracer</u> uptake

 Pretty much the same as on the previous slide, BUT in addition, nematode autofluorescence greatly hampers proper quantification of ingested particles

 Any preservation method can lead to gut evacuation and several preservatives (e.g. glutaraldehyde) greatly add to the problem of autofluorescence

experiments



courtesy Ineke Dhondt

2. Tracer experiments

methodological issues in <u>radioactive tracer</u> experiments

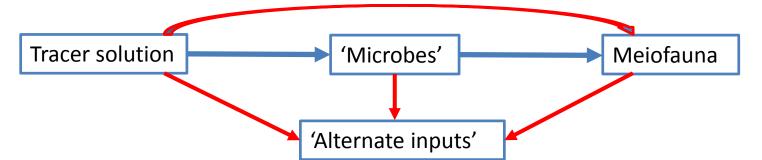
(pulse-chase) → serious risk of overestimating uptake

- extensive methodological work done by Paul Montagna, J.E. Bauer, Kevin

Carman to control for

+ alternative routes of label uptake + adsorption to body surfaces

+ homogeneous distribution of label



Elucidation of meiofaunal trophic interactions: Important results

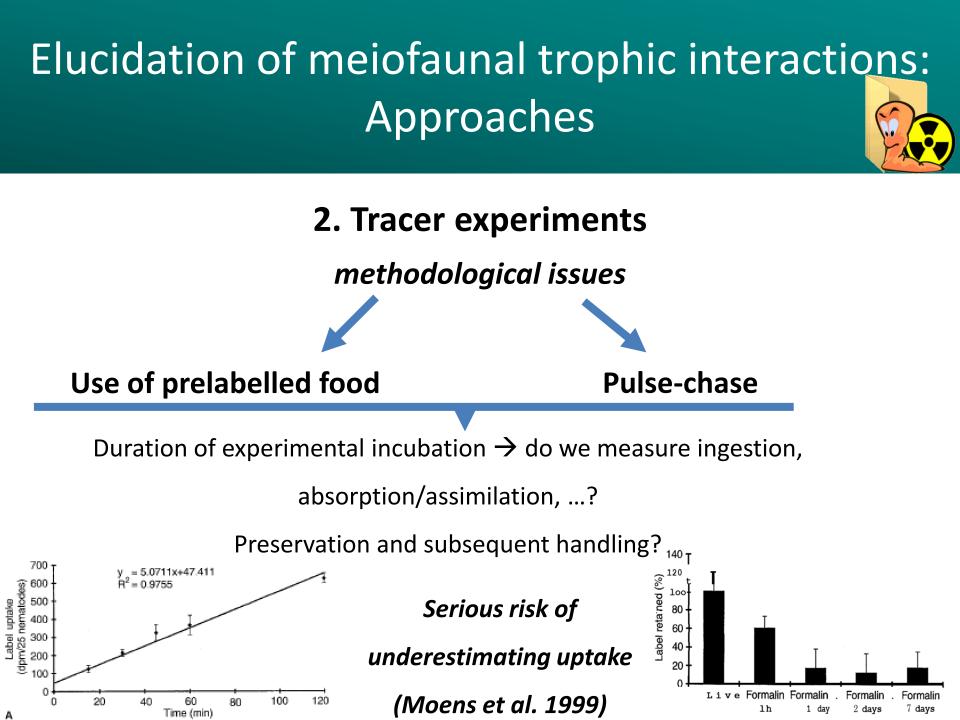
2. Radioactive tracer experiments \rightarrow <u>conclusions</u>

- Montagna (1995) 'Large variation between studies, but on average

meiofauna graze ca 1% of microbial production h⁻¹'

- Blanchard (1990), Montagna & Yoon (1991) 'Meiofaunal grazing

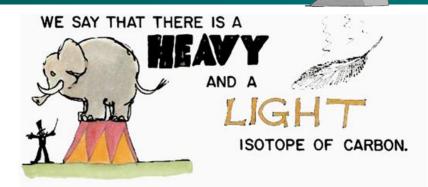
temporarily exceeds microbial production'



3. Stable isotopes

A: as tracers in enrichment exps

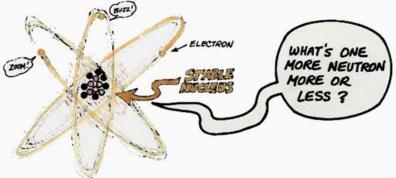
Fig. An extra neutron in the ¹³C isotope makes the nucleus more massive or "heavier" than the ¹²C isotope, but **does not affect most chemistry** that is related to reactions in the electron shell. *Fry (2008)*



¹³CARBON HAS ONE MORE NEUTRON THAN ¹² CARBON IN ITS NUCLEUS.



IN MOST CASES ¹²CARBON AND ¹³CARBON BEHAVE THE SAME BECAUSE EXTRA NEUTRONS DON'T CHANGE THE REACTIVE SPHERE OF ELECTRONS AROUND THE NUCLEUS.



in kinetic reactions, lighter isotopes usually react faster, while in exchange reactions, heavy isotopes concentrate where bonds are strongest

- → Both processes lead to isotopic fractionation, and isotopic fractionation leads to
 - different isotopic ratios between sources

H	c	lifferent isotopic ratios between consumer & resource											He				
Li	Be										B	C	N	0	F	Ne	
Na	Na Mg											A1	Si	Р	S	Cl	Ar
K	Ca	Sc	Ti	V	Cr	Mn	Fe	Со	Ni	Cu	Zn	Ga	Ge	As	Se	Br	Кſ
Rb	Sr	Y	Zr	Nb	Мо	Tc	Ru	Rh	Pđ	Ag	Cđ	In	Sn	Sb	Te	Ι	Xe
Cs	Ba	La	Hf	Ta	W	Re	Os	lr	Pt	Au	Hg	T1	Pb	Bi	Po	At	Rn
Fr	Ra	Ac															

Elucidation of meiofaunal trophic interactions: Important results

3. Stable isotopes: A: tracer experiments

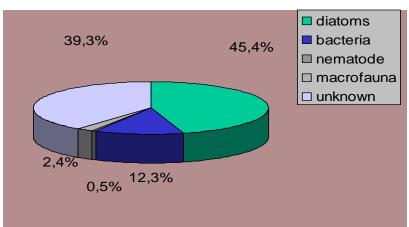
Nowadays, most radioactive tracer work has been replaced by stable isotope tracers

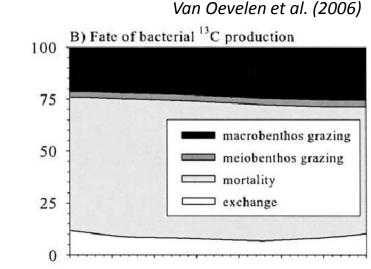
Main conclusions:

More often than not, the results indicate that meiofauna graze an insignificant

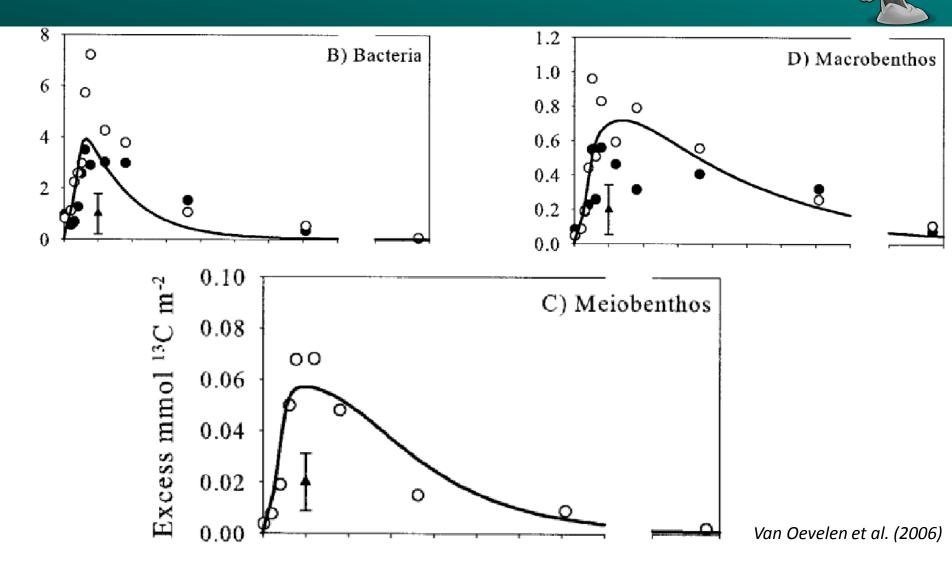
fraction of microbial production/biomass

Based on original data from Middelburg et al. (2000)





Elucidation of meiofaunal trophic interactions: Important results



Absolute quantifications: difficult and often with

conflicting results

Characterizing resource utilization and trophic

position: we are pretty much addressing the same

questions as 40 years ago

let's observe but in different ways

X. Gut content analysis in a different way: phytopigment analysis

Pioneered for meiofauna by Lidia Souza-Santos, Paulo Santos &

Jacques Castel (mid '90s)

Applied more recently for epilithic meiofauna by Nabil Majdi et

al.

Interesting enough, but...

(high biomass requirements, issues with preservations, etc...)

3B. Stable isotopes: natural abundances

A. Some possibilities

- an integrated picture of diet over the past days/weeks
- information on resources (mostly C isotopes, S would be useful but is too

'rare') → <u>you are what you eat</u>

- information on **trophic level** (cf.

trophic-level fractionation) \rightarrow mostly N

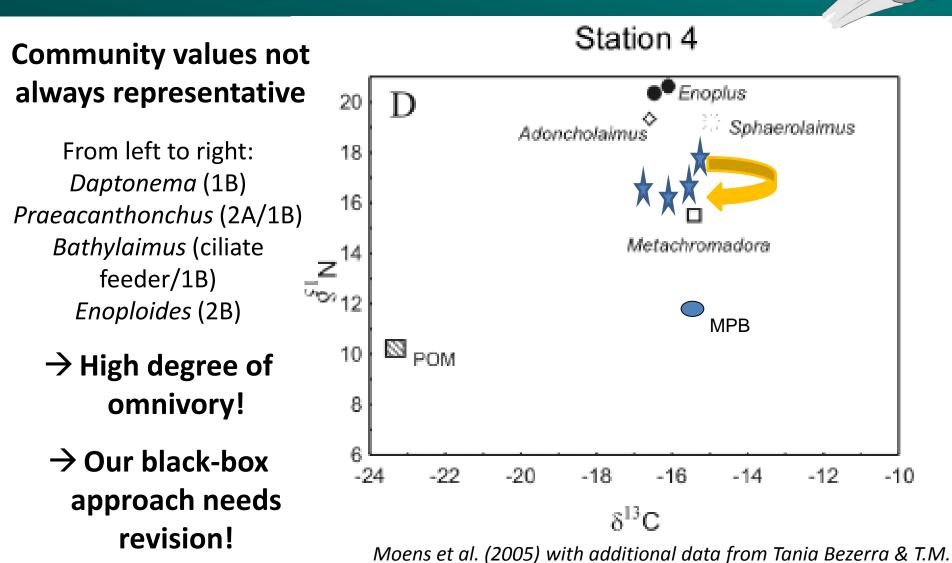
- **metrics** based on isotopes allow assessment of niche width and overlap

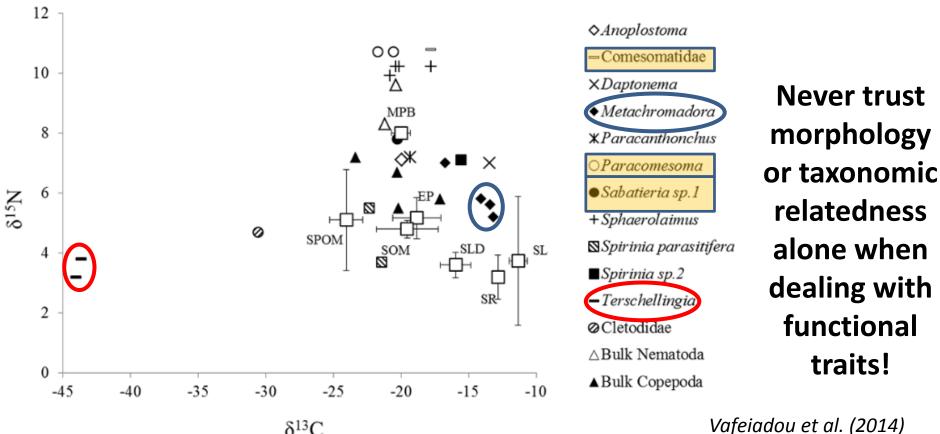
B. Some problems

only really useful if *different resources differ* enough in their isotopic

composition

- resource resolution limited
- substantial biomass (ca 5 µg of an element) required for reproducible measurements
- trophic-level fractionation appears far from constant





Vafeiadou et al. (2014)

Figure 3. Biplots of $\delta^{13}C / \delta^{15}N$ of meiobenthos from the upper 2 cm and their potential resources in seagrass beds (A) and bare sediments (B). Resource data are mean values (\pm SD) of all replicate samples per source material. Abbreviations used: SL, SR and SLD for seagrass leaves, roots and detritus, respectively; EP for epiphytes, MPB for microphytobenthos, SPOM for suspended particulate organic matter and SOM for bulk sediment organic matter.

4. Fatty acid profiles

Pioneering work on meiofauna by Daniel Leduc, Marleen De Troch and in several papers on deep-sea nematodes in group of Ann Vanreusel

Complementary info to SI

Similar limitations

Bioconversion as an additional issue

We need sufficient biomass \rightarrow pooling of many tens of inds.

interindividual variation codetermines a population's niche width and hence its environmental tolerance range (Violle et al. 2012).

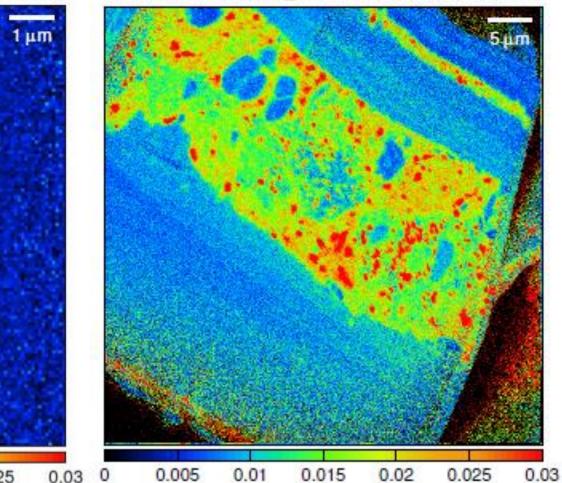
interindividual variation is key to understanding competitive interactions (both intra- and interspecific) (Violle et al. 2011) and hence community assembly and structure.

So where does that leave us?

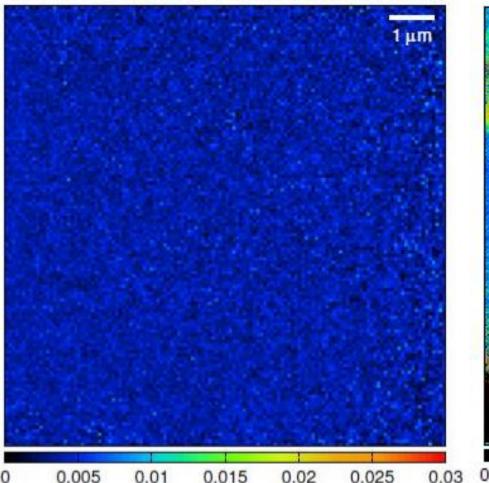
5. Stable isotope analysis in a different way:



...11-10-NEMATODE_4 : 12C15N/(12C14N+12C15N)



...11-10-NEMATODE_1 : 12C15N/(12C14N+12C15N)



5. Stable isotope analysis in a different way: NanoSIMS

A. Some possibilities

- measurements at the level of

individuals, tissues, and even single

cells

- uptake and assimilation can be

visualized

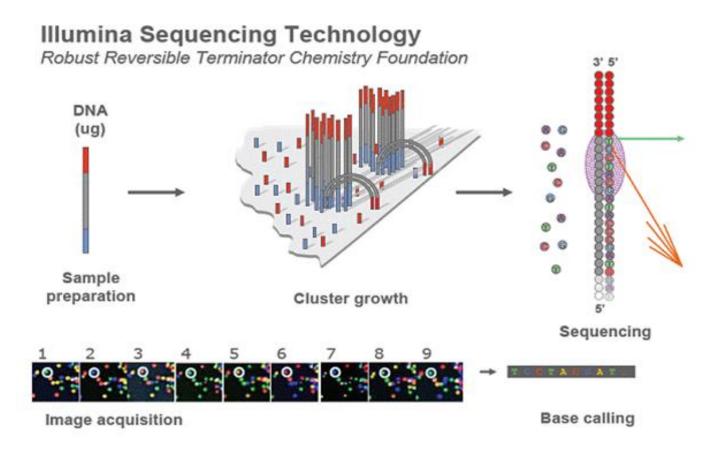
- many isotopic combinations possible

 \rightarrow S isotopes can for the first time be - used in our analyses of resource use

B. Some problems

- much specialized preparatory work
- much specialized work to analyse the data and decide on how to select the right info from the wealth of data
- extremely expensive and high-tech equipment
- analyses for the moment 20-50 times more expensive than (bulk) EA-IRMS

6. Next Generation Sequencing to analyse 'gut content'



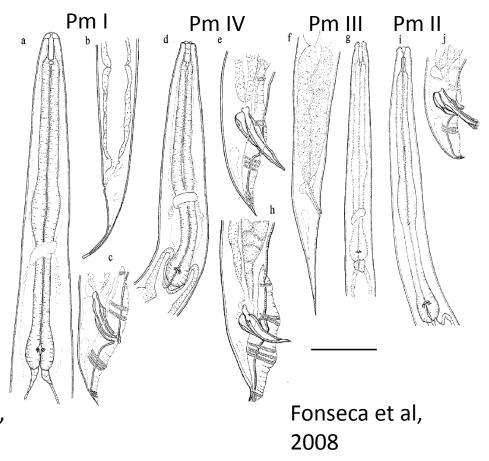
Unravelling coexistence of cryptic *Litoditis marina* species

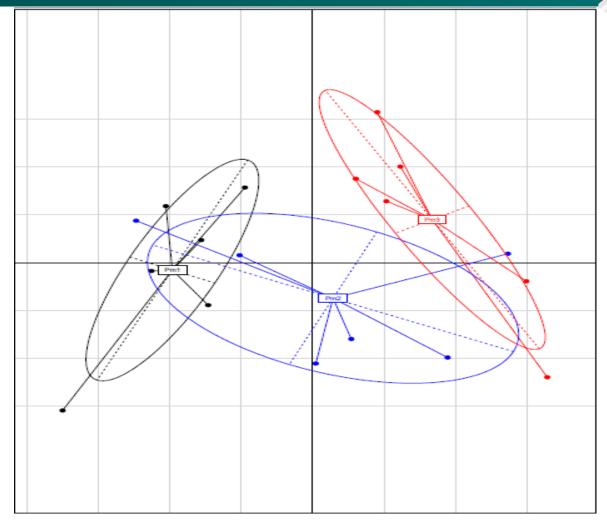
De Meester N. (2016) – PhD

Is niche-differentiation important? Is resource differentiation important in separating niches?

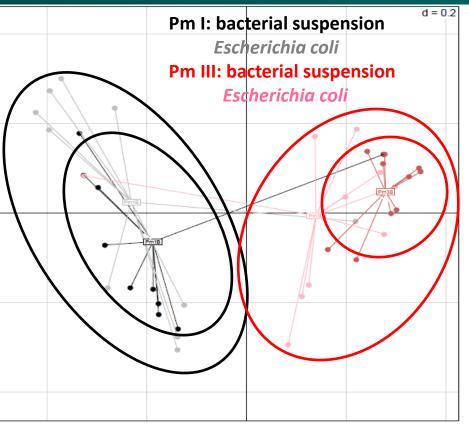
Derycke et al. (2016) – Mol Ecol

'Coexisting cryptic species of the *Litoditis marina* complex (Nematoda) have distinct microbiomes with high intraspecific variability'





Derycke et al. (2016) – Mol Ecol



Derycke et al. (2016) – Mol Ecol

Table 2 Summary of the PERMDISP and PERMANOVA statistics between the microbiomes of the four food experiment treatments (Pm1B, Pm1E, Pm3B and Pm3E) for the data set containing all OTUs and for the core OTUs. For the pairwise comparisons, significant *P*-values after Bonferroni correction are indicated in bold.

		All OTUs		Core Genome			
Food experiment		Pseudo-F	P value	Pseudo-F	P value		
PERMDISP	Species	9.04	< 0.001	7.11	0.011		
	Food	2.94	0.095	1.57	0.22		
	Species*food	6.80	< 0.001	6.65	0.001		
PERMANOVA	Species	10.97	0.001	16.56	0.001		
	Food	3.10	0.005	3.59	0.008		
	Species*food	2.02	0.049	2.46	0.043		
Pairwise test	Pm1B-Pm1E	1.65	0.236	1.62	0.13		
Pairwise test	Pm3B-Pm3E	3.98	0.004	5.50	0.001		
Pairwise test	Pm1B-Pm3B	8.78	0.004	14.71	0.001		
Pairwise test	Pm1E-Pm3E	4.81	0.004	6.1	0.002		

So we can analyse prokaryotic 'diets' of meiofauna at the level of individuals

Variability among individuals is large \rightarrow consequences at the population level?

Differences between species can clearly be analysed

We should be able to analyse eukaryotic diets in much the same way, but so far not successful

More group-specific predator-prey relationships can be analysed if suitable prey-specific primers can be developed.

Invertebrate Biology 133(2): 121–127. © 2014, The American Microscopical Society, Inc. DOI: 10.1111/ivb.12048

Diagnostic PCR can be used to illuminate meiofaunal diets and trophic relationships

Hanna Maghsoud,¹ Austin Weiss,² Julian P.S. Smith III,^{2,a} Marian K. Litvaitis,³ and Stephen R. Fegley⁴

and Smith et al. (2016) poster 76, this conference.

Elucidation of meiofaunal trophic interactions: Conclusions

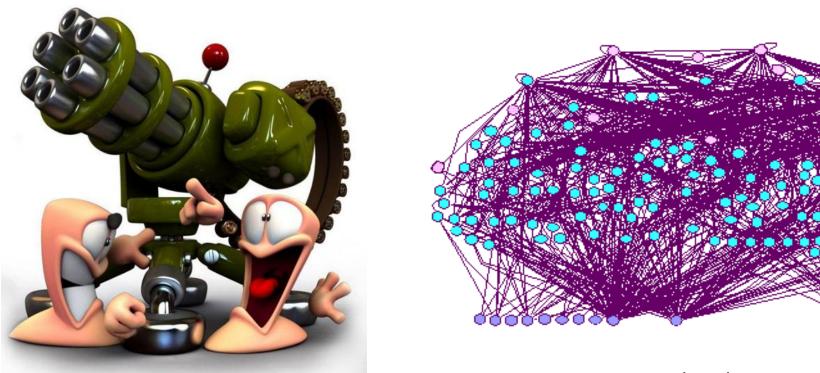
- This keynote has focused on a limited number of trophic interactions. A.o., meiofaunaciliate/flagellate and meiofauna-fungi interactions deserve more attention.
 - Despite substantial efforts and different methodological approaches, some qualitative and nearly all quantitative key questions remain under debate.
- We have to **observe** again, though with different means, before quantifying.

Elucidation of meiofaunal trophic interactions: Conclusions

- Novel technological advances open up **unprecedented opportunities** to study trophic interactions, including under natural conditions.

- They all do have their **caveats**, and some simple issues such as sample preservation effects on gut content become more pressing than ever.
- Little, if any, additional understanding on food web interactions is to be expected from analyses lumping organisms at the community, guild or family level.

Elucidation of meiofaunal trophic interactions: Conclusions



Jenny Schmid-Araya et al. (2002)



THANK YOU ALL !